

Physiology of Pgr's and Nitrogen on Crop Growth Rate, Net Assimilation Rate, Nitrate Reductase Activity and Indole Acetic Acid Oxidase Activity of Black Gram (*Vigna Mungo* L.)

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Abstract The field experiment were undertaken to study the effect of nitrogen in combination with foliar spray of bioregulators and micronutrients on growth and productivity of CO5 black gram. Photosynthetic pigments and foliage soluble protein content were estimated at different phenological phases of black gram. Seed yield were assessed at the time of harvest. Significant increase in the CGR (Crop Growth Rate) and NAR (Net Assimilation Rate) due to basal application of nitrogen 25 kg per hectare with foliar spray of urea 2% and 0.1 ppm brassinolide. The NA-ase and IAA-oxidase enzymes were also greatly altered by the basal application of nitrogen 25 kg per hectare with foliar spray of urea 2% and 0.1 ppm brassinolide treatment.

Keywords Black gram; PGR (Plant Growth Regulator); CGR (Crop Growth Rate); NAR (Net Assimilation Rate); NR-ase; IAA-oxidase; Nitrogen and yield

Introduction

Pulses are the most important crops in India because of its low cost and high quality protein. They play a major role in providing a balanced protein component in the diet of the people. Pulses contain a higher level of quality protein, nearly three times as much as cereals; therefore they are the cheapest and rich source of protein and essential amino acids and thus share a major protein of the vegetarian diet. Besides, the crops enrich the soil fertility and health in terms of addition of nitrogen and organic matter. Among pulses, black gram (*Vigna mungo* L. Hepper), occupies a unique place for its use as vegetable, and it is grown both as pure and mixed crop along with maize, cotton, sorghum and other millets. It is also known as urd bean, and it is an important pulse crop grown all over the world. It is a major component of the daily Indian diet and serves as a rich protein source (23.9%)

besides; it also contains 60.4% carbohydrates. As per the World Health Organization every man needs 80 g of pulses per day and as per the Indian Council of Medical Research, every man needs minimum consumption of 47 g of protein per day to meet requirement of the body. But at present, the per capita availability of pulses is only 30~35 g per day. Therefore, there is a need for three fold increase in pulse production as that of current production. Black gram is indeterminate in its flowering and fruiting habits and there is a competition for available assimilates between vegetative and reproductive sinks. There is limitation of source (leaves) particularly at flowering and fruiting stage. Hence, there is a need to improve LAI, CGR, NAR and LAD. Being a C₃ plant, CGR and RGR are relatively less than cereals and the major yield components are pods per plant, seeds per plant and test weight of seeds. Apart from this genetic

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makeup, the major physiological constraints limiting its production are flower drop and fruit drop (Ojeaga and Ojehomon, 1972). This performance of the crops can be overcome by foliar application of growth regulating chemicals at the crucial stages of the crop, which is one of the latest trends in agriculture. The growth regulating chemicals bioregulators can improve the LAI, SLW and SLA and play a significant role in improving the productive potential of the crop. With this above background, the present investigation was carried out.

1 Result

1.1 Crop growth rate (CGR) ($\text{g}\cdot\text{m}^{-2}\cdot\text{day}^{-1}$)

Crop Growth Rate recorded between vegetative to flowering stage (30~45 DAS) flowering to pod filling stage (45~60 DAS) and pod filling to harvest stage (60~75 DAS) is presented in (Table 1). The crop growth rate ranged from 0.54 to 0.78, 1.22 to 1.84 and 1.02 to 1.36 $\text{g}\cdot\text{m}^{-2}\cdot\text{day}^{-1}$ at 30~45, 45~60 and 60~75

DAS, respectively. The treatment T₇ (N 25 kg/ha + Urea 2% + BR 0.1 ppm) recorded the maximum CGR (0.78, 1.84 and 1.36 $\text{g}\cdot\text{m}^{-2}\cdot\text{day}^{-1}$) at all the stages of growth. There was no significant variation among the treatments at pod filling to harvest stage.

1.2 Net assimilation rate (NAR) ($\text{mg}\cdot\text{cm}^{-2}\cdot\text{day}^{-1}$)

The trend of NAR decreased from 30~45 DAS of crop growth to 60~75 DAS (Table 2). However, the declining trend was reduced by the application of nutrients and growth regulators. The NAR at various stages of crop growth viz., 30~45 DAS, 45~60 DAS and 60~75 DAS stages ranged from 0.54 $\text{mg}\cdot\text{cm}^{-2}\cdot\text{day}^{-1}$ to 0.66 $\text{mg}\cdot\text{cm}^{-2}\cdot\text{day}^{-1}$, 0.50 $\text{mg}\cdot\text{cm}^{-2}\cdot\text{day}^{-1}$ to 0.65 $\text{mg}\cdot\text{cm}^{-2}\cdot\text{day}^{-1}$ and 0.50 $\text{mg}\cdot\text{cm}^{-2}\cdot\text{day}^{-1}$ to 0.63 $\text{mg}\cdot\text{cm}^{-2}\cdot\text{day}^{-1}$ respectively. The treatment T₇ (N 25 kg/ha + Urea 2% + BR 0.1 ppm) recorded the highest NAR values at all the stages of crop growth, followed by T₃ and T₄ treatments.

Table 1 Effect of nitrogen nutrition and growth regulators on crop growth rate ($\text{g}\cdot\text{m}^{-2}\cdot\text{day}^{-1}$) in black gram at different growth stages

| Treatment | 30~45 DAS | 45~60 DAS | 60~75 DAS |
|----------------|-----------|-----------|-----------|
| T ₁ | 0.54 | 1.22 | 1.02 |
| T ₂ | 0.56 | 1.28 | 1.07 |
| T ₃ | 0.71 | 1.62 | 1.20 |
| T ₄ | 0.60 | 1.53 | 1.21 |
| T ₅ | 0.61 | 1.44 | 1.03 |
| T ₆ | 0.55 | 1.51 | 1.09 |
| T ₇ | 0.78 | 1.84 | 1.36 |
| T ₈ | 0.59 | 1.42 | 1.01 |
| T ₉ | 0.60 | 1.58 | 1.04 |
| Mean | 0.61 | 1.49 | 1.11 |
| SEd | 0.02 | 0.06 | 0.04 |
| CD (P=0.05) | 0.05 | 0.13 | 0.98 |

Note: T₁: Control; T₂: N 25 kg/ha + Urea 2% + NAA 40 ppm; T₃: N 50 kg/ha + CCC 200 ppm; T₄: N 25 kg/ha + Urea 2% + CCC 200 ppm; T₅: N 25 kg/ha + Urea 2% + Humic acid 0.1%; T₆: N 25 kg/ha + Urea 2% + Salicylic acid 100 ppm; T₇: N 25 kg/ha + Urea 2% + Brassinosteroid 0.1 ppm; T₈: N 25 kg/ha + Urea 2% + ZnSO₄ 0.5% + FeSO₄ 0.5% + Borax 0.2%; T₉: N 25 kg/ha + Water spray

1.3 Nitrate reductase activity ($\mu\text{g NO}_2\cdot\text{g}^{-1}\cdot\text{hr}^{-1}$)

The trend of NRase activity estimated at four stages of growth is presented in Table 3. Higher NRase activity was observed at the pod filling stage (60 DAS) for all the treatments including control. At the same stage, significant increase (56.2 $\mu\text{g NO}_2\cdot\text{g}^{-1}\cdot\text{hr}^{-1}$) was observed with N 25 kg/ha + urea 2% + 0.1 ppm BR (T₇) treated plants compared to other treatments. The

enzyme activity recorded for treatment T₄ was found to be statistically comparable with the values of treatment T₇.

1.4 Indole acetic acid oxidase activity (IAAO) ($\mu\text{g of unoxidised auxin}\cdot\text{g}^{-1}\cdot\text{hr}^{-1}$)

The data on IAA oxidase estimated at specific stages and expressed in terms of unoxidized auxin content is

presented in Table 4. The unoxidised auxin content was significantly increased by the foliar application of nutrients and growth regulators. Application of N 25 kg/ha + urea 2% + 0.1 ppm BR (T₇) proved to be superior (65.8, 98.5, 105.6 and 74.8 µg of unoxidised

auxin·g⁻¹·hr⁻¹) in its activity compared to all other treatments. It is also known that all the treatments were significantly superior over the control (88.1, 121.4, 147.2 and 116.0 µg of unoxidised auxin·g⁻¹·hr⁻¹) at all the stages of growth.

Table 2 Effect of nitrogen nutrition and growth regulators on net assimilation rate (mg·m⁻²·day⁻¹) in black gram at different growth stages

| Treatment | 30~45 DAS | 45~60 DAS | 60~75 DAS |
|----------------|-----------|-----------|-----------|
| T ₁ | 0.54 | 0.50 | 0.48 |
| T ₂ | 0.60 | 0.58 | 0.57 |
| T ₃ | 0.65 | 0.64 | 0.61 |
| T ₄ | 0.64 | 0.60 | 0.59 |
| T ₅ | 0.59 | 0.57 | 0.55 |
| T ₆ | 0.57 | 0.54 | 0.52 |
| T ₇ | 0.66 | 0.65 | 0.63 |
| T ₈ | 0.62 | 0.59 | 0.56 |
| T ₉ | 0.56 | 0.52 | 0.50 |
| Mean | 0.61 | 0.58 | 0.56 |
| SEd | 0.026 | 0.025 | 0.024 |
| CD (P=0.05) | 0.055 | 0.054 | 0.052 |

Note: T₁: Control; T₂: N 25 kg/ha + Urea 2% + NAA 40 ppm; T₃: N 50 kg/ha + CCC 200 ppm; T₄: N 25 kg/ha + Urea 2% + CCC 200 ppm; T₅: N 25 kg/ha + Urea 2% + Humic acid 0.1%; T₆: N 25 k/ha + Urea 2% + Salicylic acid 100 ppm; T₇: N 25 kg/ha + Urea 2% + Brassinosteriod 0.1 ppm; T₈: N 25 kg/ha + Urea 2% + ZnSO₄ 0.5% + FeSO₄ 0.5% + Borax 0.2%; T₉: N 25 kg/ha + Water spray

Table 3 Effect of nitrogen nutrition and growth regulators on NRase activity (µg NO₂·g⁻¹·hr⁻¹) in black gram at different growth stages

| Treatments | 30 DAS | 45 DAS | 60 DAS | Harvest stage |
|----------------|--------|--------|--------|---------------|
| T ₁ | 14.4 | 19.4 | 28.1 | 9.8 |
| T ₂ | 16.2 | 22.8 | 41.7 | 11.9 |
| T ₃ | 18.3 | 28.6 | 49.8 | 10.6 |
| T ₄ | 19.9 | 34.6 | 52.8 | 17.3 |
| T ₅ | 16.7 | 22.9 | 42.8 | 10.7 |
| T ₆ | 17.6 | 26.4 | 46.5 | 10.8 |
| T ₇ | 21.2 | 36.3 | 56.2 | 18.3 |
| T ₈ | 15.8 | 22.4 | 38.4 | 15.4 |
| T ₉ | 15.1 | 21.0 | 30.4 | 11.6 |
| Mean | 17.21 | 26.09 | 43.58 | 12.91 |
| SEd | 0.72 | 1.10 | 1.82 | 0.54 |
| CD (0.05) | 1.54 | 2.34 | 3.87 | 1.15 |

Note: T₁: Control; T₂: N 25 kg/ha + Urea 2% + NAA 40 ppm; T₃: N 50 kg/ha + CCC 200 ppm; T₄: N 25 kg/ha + Urea 2% + CCC 200 ppm; T₅: N 25 kg/ha + Urea 2% + Humic acid 0.1%; T₆: N 25 k/ha + Urea 2% + Salicylic acid 100 ppm; T₇: N 25 kg/ha + Urea 2% + Brassinosteriod 0.1 ppm; T₈: N 25 kg/ha + Urea 2% + ZnSO₄ 0.5% + FeSO₄ 0.5% + Borax 0.2%; T₉: N 25 kg/ha + Water spray

Table 4 Effect of nitrogen nutrition and growth regulators on IAA-oxidase in black gram at different growth stages ($\mu\text{g unoxidised auxin}\cdot\text{g}^{-1}\cdot\text{hr}^{-1}$)

| Treatments | 30 DAS | 45 DAS | 60 DAS | Harvest stage |
|----------------|--------|--------|--------|---------------|
| T ₁ | 88.1 | 121.4 | 147.2 | 116.0 |
| T ₂ | 71.3 | 118.7 | 128.6 | 78.4 |
| T ₃ | 69.0 | 109.1 | 114.9 | 89.3 |
| T ₄ | 68.7 | 101.5 | 109.4 | 78.6 |
| T ₅ | 73.8 | 116.9 | 124.3 | 84.2 |
| T ₆ | 80.7 | 111.0 | 120.4 | 92.5 |
| T ₇ | 65.8 | 98.5 | 105.6 | 74.8 |
| T ₈ | 78.4 | 105.2 | 117.6 | 86.4 |
| T ₉ | 85.6 | 120.7 | 138.5 | 109.5 |
| Mean | 75.54 | 111.22 | 122.67 | 89.75 |
| SEd | 3.33 | 4.83 | 5.42 | 4.05 |
| CD (0.05) | 7.07 | 10.24 | 11.50 | 8.58 |

Note: T₁: Control; T₂: N 25 kg/ha + Urea 2% + NAA 40 ppm; T₃: N 50 kg/ha + CCC 200 ppm; T₄: N 25 kg/ha + Urea 2% + CCC 200 ppm; T₅: N 25 kg/ha + Urea 2% + Humic acid 0.1%; T₆: N 25 kg/ha + Urea 2% + Salicylic acid 100 ppm; T₇: N 25 kg/ha + Urea 2% + Brassinosteroid 0.1 ppm; T₈: N 25 kg/ha + Urea 2% + ZnSO₄ 0.5% + FeSO₄ 0.5% + Borax 0.2%; T₉: N 25 kg/ha + Water spray

2 Discussion

Crop production is determined by crop growth rate as a function of light interception by the leaf area of a crop (Whigham, 1983). Shibles and Weber (1966) also stated that as CGR is a linear function of intercepted irradiance, maintaining higher LAI has a positive effect for higher dry matter production due to increased CGR. As observed in the present study, CGR of black gram exhibited an increasing trend from vegetative to pod filling stage followed by a decline thereafter. Since CGR is the product of NAR and LAI, this trend indicated that in black gram CGR is closely related to LAI rather than NAR. In supporting this finding, Shibles and Weber (1966) observed a strong positive correlation between CGR and LAI in soybean. Plant growth hormones played a significant role on GRR as observed in the present study. BR 0.1 ppm + Urea 2% spray in addition to recommended dose of N (25 kg/ha) strongly influenced CGR with more than 50 per cent increase over control, particularly between flowering and pod formation stages. This finding is in close confirmity with the results of Prakash et al. (2007) in sesamum. Umadevi (1998) also quoted the significant effect of BR in improving CGR in sesamum. Cycocel (200 ppm) spray in addition to N (50 kg/ha) also showed remarkable enhancement in CGR with about 30% increase over control in black

gram of the present study. This finding was corroborated with the result of Vijayakumar and Abdulkhadar (1997) in cassava. This enhancement in CGR was due to fast development of the sources as well as sink.

The net assimilation rate is a measure of net photosynthesis of leaves in crop community. In the present investigation, net assimilation rate slightly declined from vegetative to final stage of the crop. Watson (1958) suggested that the cause for the decrease in NAR with increased leaf area in crop plants was due to mutual shading of the leaves, which would decrease the leaf photosynthesis. Briggs et al. (1920), however, viewed that the decreased NAR with age was due to losses in respiration in all parts of the plants. NAR was significantly influenced by hormonal manipulation, particularly brassinolide. BR (0.1 ppm) along with Urea (2%) increased the assimilation rate by 22% during vegetative to flowering stage and 30% during flowering to pod filling stages over control. Similarly, N (50 kg/ha) + CCC (200 ppm) spray resulted in 20% and 28% improvement in NAR over control in these two stages respectively. These findings were in confirmity with the results of Kalita et al. (1994). Nemchenko et al. (1981) demonstrated the positive rate of CCC in increasing net assimilation rate of potato. Baghal and Yadhav (1992) explained

the significant role of BR in enhancing the NAR in black gram besides arriving strong correlation with seed yield. The increase in NAR due to BR could be assigned to its growth regulating activity, since its activity is analogous to auxin and gibberellins.

The nitrate reductase is the rate-limiting enzyme in nitrogen assimilation and is a key point of metabolic regulation (Eilrich and Hageman, 1973) in crop plants. Thus, NRase is intimately associated with the plant growth and development (Sinha and Nicholas, 1981). While observing the pattern of NRase activity in groundnut throughout the growth period, Sung and Sun (1990) reported that NRase activity declined rapidly during post flowering stages. However in the pre-seed foliage. The enzyme activity was further triggered by combined effect of BR. Similar to this finding, Sairam (1994) study high NRase activity was extended up to pod filling stage, which might be due to the additional dose of Urea applied through observed a significant increase in NRase activity due to brassinolide application in wheat. Jaisingh et al. (1993) also recorded an increased rate of enzyme activity due to brassinolide application in *Cicer aurietinum*. Besides brassinolide, CCC was also found effective in triggering this enzyme activity as revealed in the present study. Wasnik and Bagga (1992) also observed the significant effect of CCC in enhancing the enzyme activity in mungbean.

The IAA oxidase activity determines the auxin levels and thereby apical dominance. It was also reported that IAA oxidase activity was low in region of high auxin content and high in region of low auxin content (Galston and Dalberg, 1954). In the present investigation, the time trend of IAA oxidase activity revealed a decreasing trend up to pod filling stage. Hormonal treatments played their differential role in altering the auxin level. The treatment combination with brassinolide effectively suppressed the enzyme activity and maintained higher level of auxin in leaves. This finding was in close confirmity with the result of Han et al. (1988), in which foliar application of brassinosteroid increased IAA synthesis in tobacco. Helmy et al. (1997) also had similar finding with the view that brassinosteroid application at 30 mg/L had enhanced the auxin content in broad bean. Bindu

Joseph (2000) also revealed the significant influence of BR on lowering the activity of IAA oxidase in groundnut up to flowering stage of the crop. Thus increase in auxin content by BR application would have contributed to increased reproductive growth. BR acted synergistically with auxin in stimulating cell elongation (Katsumi, 1985) and ethylene production (Arteca et al., 1995) suggesting that effects of BR are mediated through auxin (Takeno and pharis, 1982) or that BRs enhance tissue sensitivity to auxin (Mandava, 1988). BR induced increase in fresh weight of squash cotyledons was accompanied by an increase in auxin and decrease in ABA levels (Eun et al., 1989).

3 Materials and Methods

The present investigation was undertaken under field condition to study the effect of nutrients and plant growth regulators on growth and productivity of black gram variety CO5. The research experiment was carried out at Millet Breeding Station, Tamil Nadu Agricultural University, Coimbatore during July to October, 2007. Growth regulators like Naphthalene Acetic Acid (NAA), Salicylic Acid (SA), Cycocel (CCC), Brassinosteroid (BR), Humic Acid and the nutrients such as Nitrogen, DAP, Boric Acid, Ferrous Sulphate, Zinc Sulphate were used. The data were statistically analyzed with the Design of Randomized Block Design with three replication and the Plot size of 4 × 3 m with Spacing of 30 × 10 cm. In this research study has nine treatments and the details are T₁: Control; T₂: N 25 kg/ha + Urea 2% + NAA 40 ppm, T₃: N 50 kg/ha + CCC 200 ppm; T₄: N 25 kg/ha + Urea 2% + CCC 200 ppm; T₅: N 25 kg/ha + Urea 2% + Humic acid 0.1%; T₆: N 25 kg/ha + Urea 2% + Salicylic acid 100 ppm; T₇: N 25 kg/ha + Urea 2% + Brassinosteroid 0.1 ppm; T₈: N 25 kg/ha + Urea 2% + ZnSO₄ 0.5% + FeSO₄ 0.5% + Borax 0.2%; T₉: N 25 kg/ha + Water spray. The CGR was calculated by employing the formula of Watson (1956) and expressed as g·m⁻²·day⁻¹. NAR (Net Assimilation Rate) was calculated by using the formula of Gregory et al. (1917) and subsequently modified by Williams (1946) and expressed as mg·cm⁻²·day⁻¹. Employing the formula of Nicholas et al. (1976), the NR'ase enzyme activity was calculated and expressed as μg NO₂-g⁻¹·hr⁻¹. The IAA-oxidase enzyme activity was

estimated by using the formula of Parthasarathy et al. (1970) and the enzyme activity was expressed as μg of unoxidised auxin $\cdot\text{g}^{-1}\cdot\text{hr}^{-1}$.

References

- Arteca R.N., 1995, Brassinosteroids, In: Davies P.J. (ed.), Plant Hormones Physiology, biochemistry and molecular biology, Kluwer Academic Press, Dordrecht, The Netherlands, pp.206-213
- Baghal M.S., and Yadav H.S., 1992, Response of blackgram to dates of sowing and growth regulators to physiological parameters and seed yield, *Bharatiya Krishi Anusandhan Patrika*, 9: 38-42
- Bindu Joseph, 2000, Physiological and biochemical effects of brassinolide on the productivity of groundnut, M.Sc (Ag.) thesis, Tamil Nadu Agricultural University, Coimbatore
- Briggs G.E., F. Kidd, and C. West, 1920, A quantitative analysis of plant growth, *Pt. Int. Ann. Appli Biol.*, 7: 103 <http://dx.doi.org/10.1111/j.1744-7348.1920.tb05107.x>
- Eilrich G.L., and R.H. Hageman, 1973, Nitrate reductase activity and its relationship to vegetative and grain nitrogen in wheat (*Triticum aestivum* L.), *Crop Sci.*, 13(1): 59-66 <http://dx.doi.org/10.2135/cropsci1973.0011183X001300010018x>
- Eun J.S, S. Kurashi, and N. Sakurai, 1989, Changes in levels of auxin and abscisic acid and the evolution of ethylene in squash hypocotyls after treatment with brassinolide, *Plant Cell Physiol.*, 30(6): 807-810
- Galston A.W., and Dalberg L.Y., 1954, The adaptive formation and physiological significance of Indole acetic acid oxidase, *Amer. J. Bot.*, 41(5): 373-389 <http://dx.doi.org/10.2307/2438724>
- Gregory P.J, Kidd G.E., and West C., 1917, Growth analysis, In: Noggle G.R., and Fritz (eds.), *Introductory Plant Physiology*, Prentice Hall, New Delhi, pp.586-588
- Helmy Y.I., Sawan O.M.M., and Abdel Halim S.M., 1997, Growth, yield and endogenous hormones of broad bean plants as affected by brassinosteroids, *Egyptian J. Hort.*, 24(1): 109-115
- Jai S., Nakamura S., and Ota Y., 1993. Effect of epi-brassinolide on gram (*Cicer arietinum*) plant grown under water stress in juvenile stage, *Indian J. Agric. Sci.*, 63(7): 395-397
- Kalita P., Dey S.C., Chandra K., and Upadhaya L.P., 1994, Effect of foliar application of N on morphological traits on pea (*Pisum sativum*), *Indian J. Agric. Sci.*, 64: 850-852
- Katsumi M., 1985, Interaction of brassinosteroids with IAA and GA3 in the cell elongation of cucumber hypocotyls sections, *Plant Cell Physiol.*, 26: 615-625
- Mandava M.B., 1988, Plant growth providing brassinosteroid, *Ann. Rev. Pl. Physiol. and Mol. Biol.*, 39: 23-32 <http://dx.doi.org/10.1146/annurev.pp.39.060188.000323>
- Nemchenko V.V., Yu. A., Ver Shinin and Khalitor A., 1981, Effectiveness of chloro choline chloride applied to potatoes, *Khimiya vsali Kom Khogyaistre*, 9: 42-45
- Nicholas J.C., Harper J.E., and Hageman R.H., 1976, Nitrate reductase activity in soybean I. Effect of light and temperature, *Plant Physiol.*, 58(6): 731-735 <http://dx.doi.org/10.1104/pp.58.6.731> PMID:16659755 PMCID:PMC542297
- Ojeaga O.O., and Ojehomon, 1972, Fruit abscission in cowpea (*Vigna unguiculata* (L) wasp.), *J.Exp.Bot.*, 23: 751-761 <http://dx.doi.org/10.1093/jxb/23.3.751>
- Parthasarathy K., Balua D.R.C., and Rao P.S., 1970, Studies on sandal spur. VIII polyphenoloxidase activity and metabolism of sandal (*Santalum album*) in healthy and diseased plant, *Proc. Indian Acad. Sci.*, 72: 277-284
- Prakash M., 2007, Effect of 28- homobrassinolide on morphological and yield parameters of sesame, *Indian J. Plant Physiol*, 12(1): 91-94
- Sairam R.K., 1994, Effect of homobrassinolide application on plant metabolism and grain yield under irrigated and moisture stress conditions of two wheat varieties, *Plant Growth Regul.*, 14(2): 173-181 <http://dx.doi.org/10.1007/BF00025220>
- Shibles R.M., and Weber C.R., 1965, Interception of solar radiation and dry matter production by various soybean planting pattern, *Crop Sci.*, 6(1): 49-52
- Sinha S.K., and Nicholas J.D., 1981, Nitrate reductase in relation to drought, In: Palag L.G., and Aspinall D. (eds.), *Physiology and Biochemistry of Drought Resistance*, Academic Press, Sydney
- Sung F.J.M., and Sun Y.W., 1990, Seasonal patterns of nitrate reductase and nitrogenase activities in *Arachis hypogaea*, *Field Crops Res.*, 25(3-4): 215-222 [http://dx.doi.org/10.1016/0378-4290\(90\)90004-U](http://dx.doi.org/10.1016/0378-4290(90)90004-U)
- Takeno K., and Pharis R.P., 1982, Brassinosteroids induced bending of leaf lamina of dwarf rice seedlings: An auxin mediated phenomenon, *Plant Cell Physiol.*, 23(7): 1275-1281
- Umadevi T.K., 1998, Studies on physiological basis of partitioning and its regulation for maximizing yield in Sesamum (*Sesamum indicum* L.), M.Sc. (Ag.) Thesis, Tamil Nadu Agricultural University, Coimbatore
- Vijayakumar M., Abdulkhadar J.B.M., 1997, Physiological parameters of cassava influenced by ethrel and cycocel, *Madras Agric. J.*, 84(8): 509-510

- Wasnik K.G., and Bagga A.K., 1992, Effects of CCC on nitrate reductase activity and grain yield in mung bean (*Vigna radiata*), Indian J. Plant Physiol., 35(1):104-107
- Watson D.J. 1958, The dependence of net assimilation rate on leaf area index, Ann. Bot., 23: 37-54
- Watson D.J. 1956, The dependence of net assimilation rate on leaf area index, Ann. Bot., 18: 29-31
- Whigham D.K., 1983, Soybean, In: Smith W.H. (ed.), Symposium on potential productivity of field crops under different environment, IRRI, Los Banos, pp.205-225
- Williams S.R.F., 1946, Methods of growth analysis, In: Sestak Z.J., Catasky and Jouris P.J. (eds.), Plant photosynthetic production manual and methods, Drow, Jenk, N.U. Publishers, The Hague, pp.348-391